

Effect of Amino Acid Application on Physiological and Biochemical Properties of Salt-Stressed Broccoli Seedlings

The effect of exogenous amino acid on some physiological and biochemical properties of broccoli seedlings under salt stress was investigated. Two products containing amino acids (Ga and Pr) were applied from the soil to the root zone of the plant 3 times with one-week intervals in this study. The solutions prepared with 0 and 100 mM NaCl were applied to the plant as irrigation water. In the study, the effects of salt stress and applications on H₂O₂, MDA, proline sucrose, catalase (CAT), superoxide dismutase (SOD), indole acetic acid (IAA), salicylic acid (SA), gibberellic acid (GA) and abscisic acid (ABA) content of seedlings were investigated. The content of H₂O₂, MDA, proline, sucrose, CAT, SOD and ABA increased, while the content of IAA, GA and SA in the plant decreased with salinity. However, with exogenous amino acid applications, the effect of salt stress on these parameters in the plant was alleviated, thus contributing to the increase in the tolerance of broccoli seedlings to salt stress.

Keywords: Broccoli, antioxidant enzyme, hormone, salinity

Introduction

Salinity stress is one of the major abiotic stresses threatening agricultural production worldwide. About 20% of the irrigated agricultural lands in the world have salinity problems. It is estimated that the area affected by salinity will be approximately 50% of the total agricultural land by 2050 (Kumar et al., 2020; Zhao et al., 2021).

With salinity, damages occur in plants in terms of various physiological, biochemical and molecular properties, and plant growth is adversely affected. Salt stress reveals its effect on the plant with osmotic and ion stress. With salt stress, high concentrations of Na⁺ accumulate in plant cells, reaching toxic levels and causing disruption of ion homeostasis. Ion imbalance and water deficiency in plant cells under salt stress cause osmotic stress, resulting in many changes such as decrease in cell turgor pressure, deterioration of the structure of the plasma membrane (Park et al., 2016). Salinity reveals these stresses by reducing plant water and nutrient uptake. Salt stress also causes various physiological and molecular changes together, suppresses cell division and expansion, inhibits photosynthesis and reduces plant growth. The activities of enzymes such as ribulose-1,5-bisphosphate carboxylase/oxygenase (RuBisCO), which are effective in photosynthesis, or protein stability are affected by salinity, in addition, salinity changes the levels of sugars such as sucrose, fructose and glycolysis in the plant (Zhu et al., 2002; Zhao et al., 2021). Salinity also disrupts various physiological and biochemical processes such as reactive oxygen species (ROS) (such as O₂, H₂O₂, O₂^{•-} and OH[•]) formation and membrane leakage (Mushtaq et al., 2020).

1 Broccoli is known to be tolerant to moderate salinity. It has been stated
2 that salt stress causes a decrease in growth with a two-stage effect in the roots
3 and salt accumulated in the leaves of broccoli. However, the response to salt
4 stress differs depending on the level of salt stress and the cultivars of broccoli
5 (Chevilly et al., 2021).

6 There are various applications to mitigate of salt stress, which causes
7 significant damage to plants. One of them is the application of amino acids. It
8 is known that salt stress creates various changes in the plant with its effects on
9 the amino acid content, which is also involved in plant metabolism. For this
10 reason, researchers have obtained results that the damage caused by stress on
11 the plant can be alleviated by applying amino acids exogenous to the plant.
12 Amino acids regulate ion transport in the plant, modulate stomatal opening,
13 affect the synthesis and activity of some enzymes, gene expression and redox
14 homeostasis (Rai, 2002). In previous studies, important effects of amino acid
15 applications against various abiotic stress factors, including salt stress, were
16 determined (Wang et al., 2017; Haghghi et al., 2020; Matysiak et al., 2020;
17 Peña Calzada et al., 2022).

18 The aim of this study is to determine the effect of exogenous amino acid
19 application on some physiological and biochemical properties of broccoli
20 seedlings under salt stress.

23 **Materials and Methods**

25 *Plant Material and Experimental Design*

27 A pot study was carried out in the controlled greenhouse conditions. To
28 obtain seedlings, the broccoli (*Brassica oleracea* var. *italica*) seeds were sown
29 in viols containing peat:perlite (2:1). Seedlings with about 2-3 leaves were
30 planted in 1.3 liter pots in a mixture of soil, peat, sand and manure (2:1:1:1).
31 After planting the seedlings, 30 ml of amino acid solution (Ga: Gluten amin
32 and Pr: Protein) was drenched to each plant root zone.

33 Both of these products contain aspartic acid, gamma-aminobutyric acid,
34 glutamic acid, alanine, arginine, phenylalanine, glycine, hydroxyproline,
35 histidine, isoleucine, leucine, lysine, methionine, proline, serine, tyrosine,
36 threonine, tryptophan and valine (Ga total %45 and Pr total 41%) in different
37 ratios.

38 The amino acid treatments were repeated three times at one-week
39 intervals. Salinity application was started with irrigations after the first amino
40 acid application. The solutions prepared with 0 and 100 mM NaCl were
41 applied to the plants as irrigation water. Experiments were conducted with
42 randomized plots design: a total of 108 plants were used with three replications
43 and 6 plants per repeat. The experiment was terminated 40 days after the
44 seedling planting. Some physiological and biochemical analyses were made.

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Hydrogen peroxide (H₂O₂), Malondialdehyde, Sucrose and Proline

H₂O₂ and MDA content of leaf tissues were determined according to method of Liu et al. (2014). The MDA content was determined by spectrophotometrically at 532 and 600 nm absorbance (Sahin et al., 2018). H₂O₂ was determined at 390 nm in spectrophotometer (Sahin et al., 2018).

The content of sucrose in the samples was determined in the spectrophotometer at a wavelength of 620 nm (Wu et al., 2011). Proline extraction and proline content were determined according to method of Bates et al. (1973) and the samples were measured at 520 nm with spectrophotometer.

CAT and SOD Enzyme Activities

Fresh leaf samples were homogenized in the extraction solution according to the method specified by Angelini et al. (1990) and Angelini and Federico (1989) and the obtained supernatant was used to determine enzyme activities. CAT activity was determined by the decrease in absorbance of H₂O₂ at 240 nm. SOD activity at 560 nm by spectrophotometrically (Liu et al., 2014).

Hormone Content

Extraction and purification processes were performed as described by Battal and Tileklioglu (2001) and Kuraishi et al. (1991). The hormones were determined by HPLC using a Zorbax Eclipse-AAA C-18 column (Agilent 1200 HPLC). Abscisic acid (ABA), gibberellic acid (GA), indole acetic acid (IAA) and salicylic acid (SA) were defined at 265 nm with a UV detector (Turan et al., 2014).

Statistical Analysis

A two-way ANOVA was used for data analysis and means comparison was made according to the Duncan multiple comparison test using SPSS program.

Results and Discussion

The effects of salt stress and amino acid application on H₂O₂, MDA, proline, sucrose, CAT, SOD, IAA, GA, SA and ABA content of broccoli seedlings are given in Figure 1, 2, 3, 4, 5 and 6.

With salinity, the amount of H₂O₂, MDA (Figure 1), proline (Figure 2), sucrose (Figure 3), CAT, SOD (Figure 4) and ABA (Figure 6) increased significantly, while the content of IAA, GA and SA (Figure 5) decreased. The increase in plant H₂O₂ and MDA content with salinity was quite high compared to the control. The amount of proline, sucrose, CAT, SOD and ABA increased

1 by 95%, 26%, 207%, 324% and 224%, respectively, compared to the control in
2 plants under salt stress without treatment. The content of IAA, GA and SA
3 decreased with salinity by 26%, 43% and 32%, respectively. However, the
4 effects of exogenous amino acid applications to plants and salinity in terms of
5 these parameters have changed. When plants in salty conditions are compared
6 with each other; H₂O₂, MDA, proline, CAT, SOD and ABA contents of treated
7 (Pr and Ga) plants were lower by 64-52%, 71-40%, 60-47%, 7-40%, 2-37%
8 and 30-44% (respectively) than control plants under the same conditions. On
9 the other hand, sucrose, IAA, GA and SA contents treatments (Pr and Ga) were
10 increased by 82-113%, 25-259%, 95-121% and 61-30% (respectively)
11 compared to control in salty conditions.

12 It was determined that H₂O₂, MDA, proline, sucrose, CAT and SOD
13 increased in broccoli seedlings with salinity, in this study. Thus, the plant
14 started to react to salt and damage symptoms appeared in plant morphological
15 features. Similarly, in another study, increased salinity in broccoli decreased
16 leaf area, shoot length, root length, shoot and root dry weights, and increased
17 antioxidant enzyme activities such as SOD, CAT and ascorbate peroxidase
18 (APX) (Ali et al., 2022). Also, according to Akram et al. (2020) stated that the
19 activities of total phenolics, H₂O₂, MDA, glycine betaine, proline, ascorbic
20 acid, CAT, SOD and peroxidase (POD) enzymes increased with increasing salt
21 stress in broccoli. With salt stress, excessive H₂O₂ production occurs in the
22 plant and it oxidizes amino acids such as methionine and cysteine from Calvin
23 cycle enzymes, causing programmed cell death (Mushtaq et al., 2020). In salt
24 stress, the plant antioxidant defense system detoxifies ROS and protects the
25 plant against oxidative damage caused by stress by providing the balance of
26 ROS formation. Excessive ROS production caused by salt stress is one of the
27 reasons that prevent morphological, physiological and biochemical activities in
28 plants by strengthening the antioxidant defense system (Hasanuzzaman et al.,
29 2021). Oxidative stress in plants occurs with enzymatic (SOD, CAT etc.) and
30 non-enzymatic (ascorbic acid, phenolic alkaloids, flavonoids, carotenoids etc.)
31 antioxidant defense mechanisms and endogenous protection mechanism
32 (Hasanuzzaman et al., 2020).

33 It is stated that various hormones (such as cytokinins, abscisic acid, auxin,
34 jasmonic acid, gibberellin and ethylene) play a role in the improvement of salt
35 stress in plants (Mushtaq et al., 2020; Raza et al., 2022; Vaishnav and
36 Chowdhury, 2023). In this study, the effect of salinity on plant hormone
37 content was significant. GA improves plant metabolism processes by
38 regulating membrane permeability, enzymatic activity, various osmolytes and
39 ion uptake in the plant, thus increasing plant tolerance to abiotic stresses
40 (Sharma et al., 2015; Raza et al., 2022). We investigated that GA content of
41 broccoli seedling showed a significant decrease with salinity. IAA is a
42 versatile phytohormone that has an active role in plant growth and
43 development, especially in stressful conditions. IAA improves the antioxidant
44 defense system and plant tolerance in the plant and thus reduces the oxidative
45 damage caused by abiotic stress (Raza et al., 2022). In this study, IAA content
46 of broccoli seedling was decreased under salt stress. SA is a phenolic

1 compound that modulates pathogenesis-related protein expression and plays a
2 role in plant growth, development and abiotic stress response as well as plant
3 defense responses. Salicylic acid helps the plant respond to and counteract
4 various abiotic stresses (Kang et al., 2014; Raza et al., 2022). SA contents
5 showed a significant decrease with salinity, in this study. ABA is a well-known
6 phytohormone with its effect and role in the adaptation of plants to various
7 abiotic stresses, and it is also called stress hormone. In response to abiotic
8 stress in the plant, endogenous ABA level rises, triggering specific signaling
9 pathways and altering gene expression levels (Danquah et al., 2014; Albacete,
10 2020; Raza et al., 2022). There was a significant increase in the ABA content
11 of broccoli seedlings with salinity in this study.

12 In this study, the effects of the above-mentioned salt stress on broccoli
13 seedlings changed significantly with amino acid treatments. The increase in
14 H₂O₂, MDA, proline, sucrose, ABA, CAT and SOD activities with salinity was
15 less with amino acid treatments. The reduction in IAA, SA and GA content
16 caused by salt stress was lower with amino acid application. Amino acids are
17 biostimulants that have a positive effect on plant growth and reduce damage
18 caused by abiotic stresses (Ali et al., 2019). Similarly, it was determined that
19 the amino acid could reduce the harmful effects of ROS and increase the
20 tolerance of wheat seedlings under salt stress conditions (Bahari et al., 2013).
21 Peña Calzada et al. (2022) determined that the amino acid mixture sprayed
22 from the leaves increases the accumulation of K⁺ in the plant under salt stress,
23 and affects biological processes such as osmolytes, photosynthetic pigments,
24 and eliminates the morphological and physiological damages caused by
25 salinity. In addition, it was stated that it caused a decrease in Na⁺ accumulation
26 and MDA concentration. The role of amino acids in abiotic stress tolerance are
27 involved with mechanisms that compatible osmolytes, regulate pH and act as
28 nitrogen and carbon reserves (Ali et al., 2019). It has been determined that
29 amino acids can alleviate the salt stress membrane by showing similar effects
30 in broccoli seedlings, thus providing tolerance against salt stress with less
31 damage to plant development.

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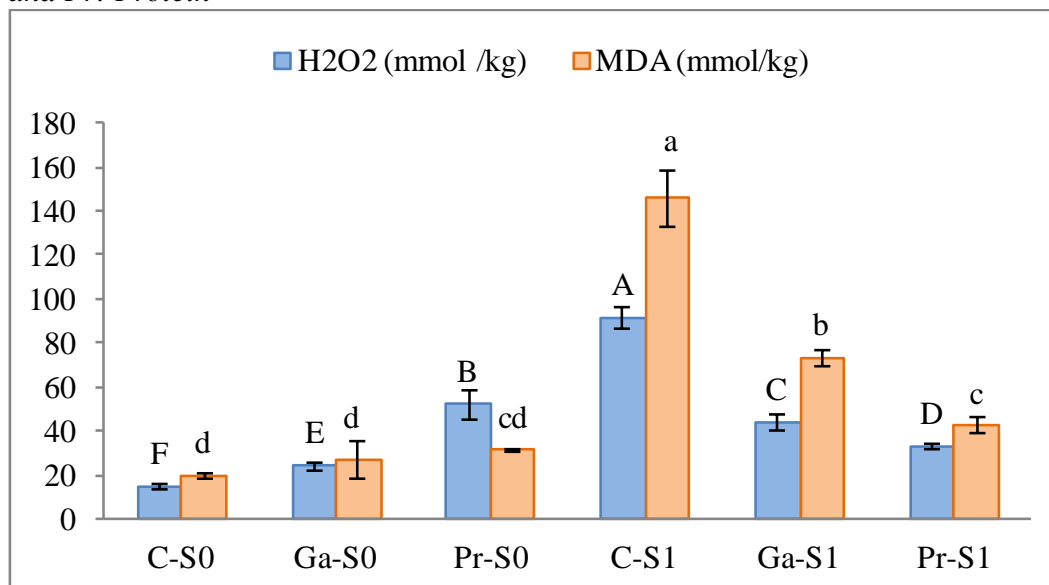
34 **Conclusion**

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36 In this study, the effect of salt stress on some physiological and
37 biochemical properties of broccoli seedlings was investigated, and significant
38 effects were observed on these properties with salt stress and thus damage to
39 the plant occurred. On the other hand, it was determined that amino acid
40 application could alleviate the damage in broccoli seedlings by changing the
41 effect of salt stress on these parameters. However, further research is needed to
42 determine the precise role of amino acids in this effect of salt stress on broccoli
43 seedlings.

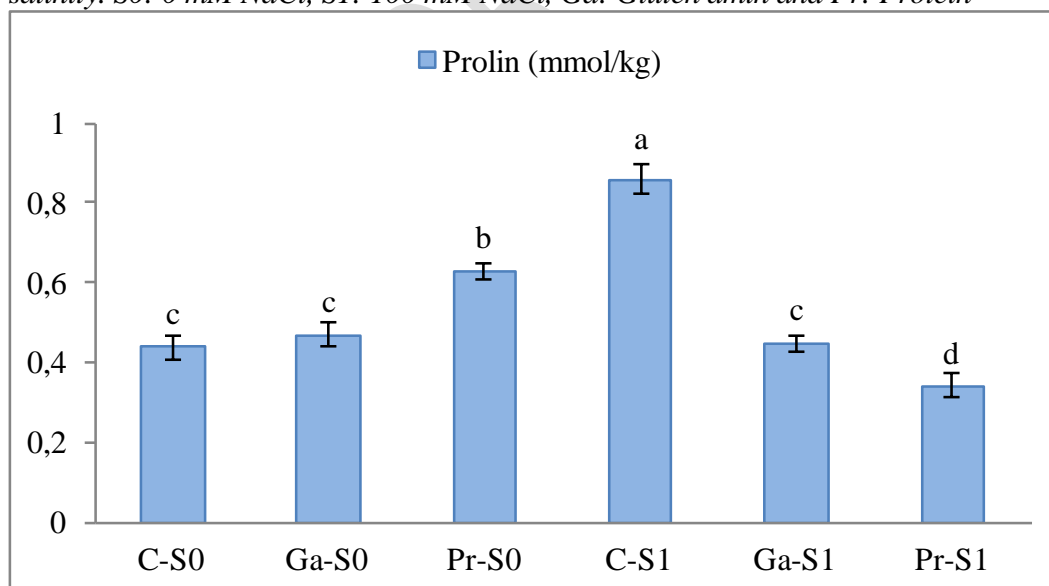
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1 **Figure 1.** The effects of amino acid treatments on H_2O_2 and MDA content of
 2 broccoli under salinity. S0: 0 mM NaCl, S1: 100 mM NaCl, Ga: Gluten amin
 3 and Pr: Protein



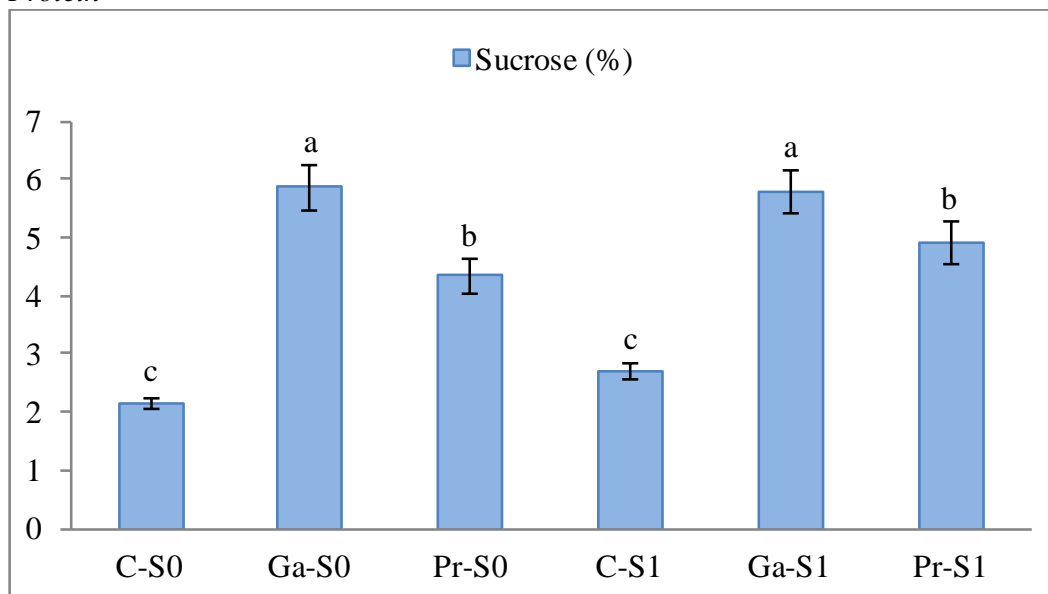
4 The difference between the means indicated by different uppercase letters and lower case
 5 letters in the same bar is statistically significant (Duncan multiple comparison test, $P < 0,001$).
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8 **Figure 2.** The effects of amino acid treatments on prolin content of broccoli under
 9 salinity. S0: 0 mM NaCl, S1: 100 mM NaCl, Ga: Gluten amin and Pr: Protein



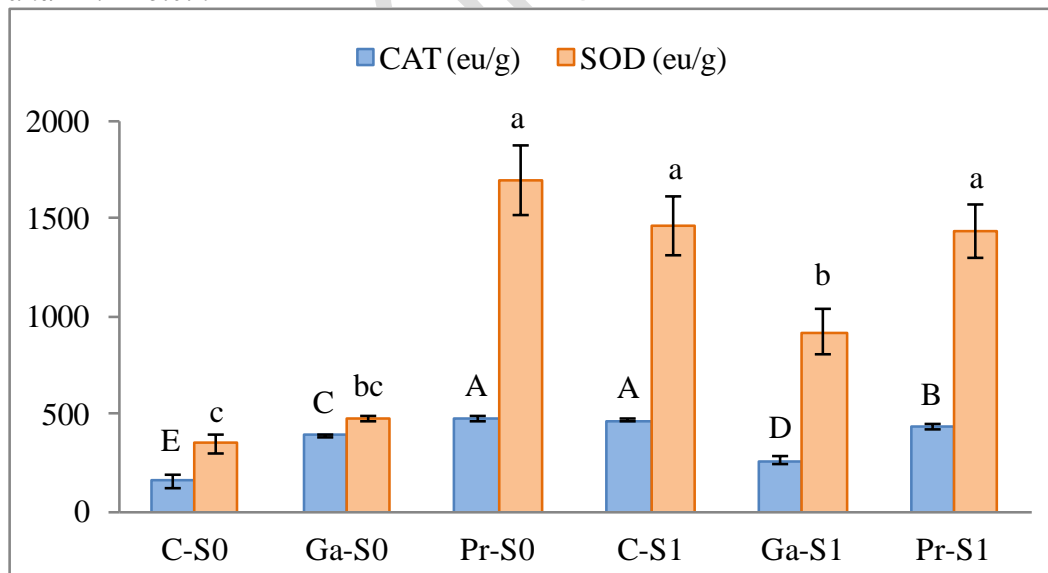
10 The difference between the means indicated by different uppercase letters and lower case
 11 letters in the same bar is statistically significant (Duncan multiple comparison test, $P < 0,001$).
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1 **Figure 3.** The effects of amino acid treatments on sucrose content of broccoli
 2 under salinity. S0: 0 mM NaCl, S1: 100 mM NaCl, Ga: Gluten amin and Pr:
 3 Protein



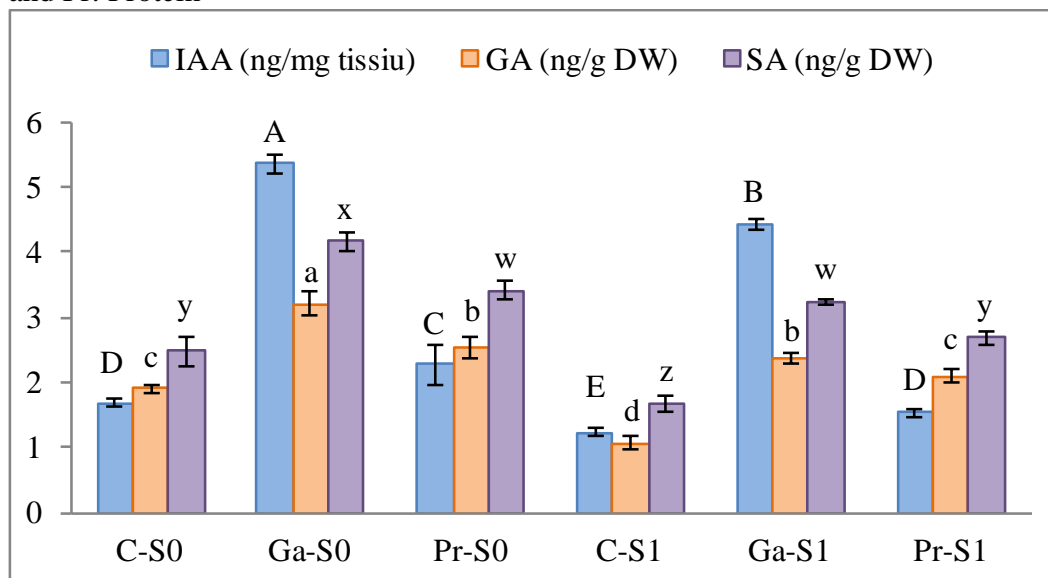
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 5 The difference between the means indicated by different uppercase letters and lower case
 6 letters in the same bar is statistically significant (Duncan multiple comparison test, P<0,001).
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8 **Figure 4.** The effects of amino acid treatments CAT and SOD activity of
 9 broccoli under salinity. S0: 0 mM NaCl, S1: 100 mM NaCl, Ga: Gluten amin
 10 and Pr: Protein



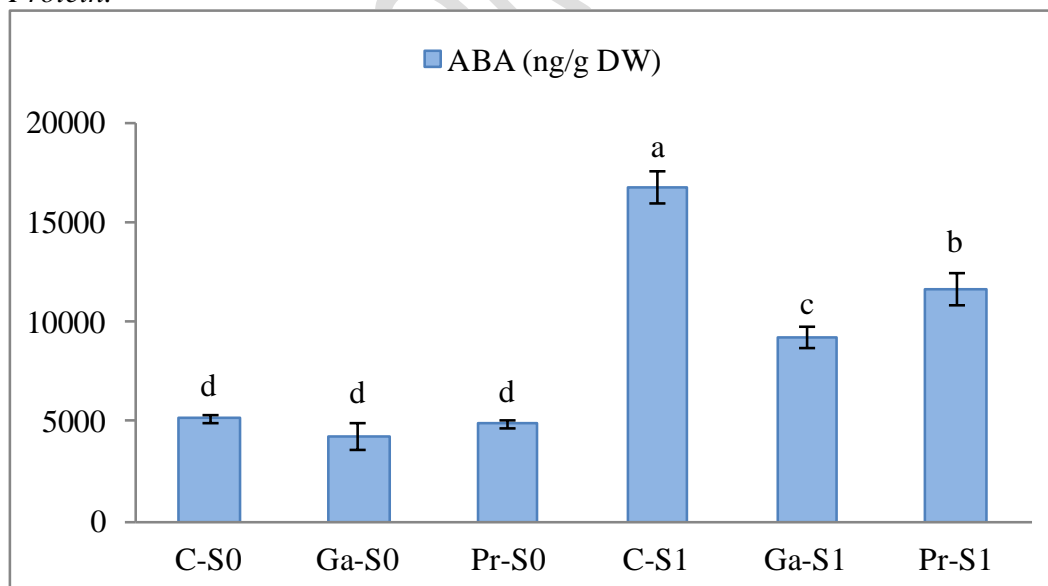
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 12 The difference between the means indicated by different uppercase letters and lower case
 13 letters in the same bar is statistically significant (Duncan multiple comparison test, P<0,001).
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1 **Figure 5.** The effects of amino acid treatments on IAA, GA and SA content of
 2 broccoli under salinity. S0: 0 mM NaCl, S1: 100 mM NaCl, Ga: Gluten amin
 3 and Pr: Protein



4
 5 The difference between the means indicated by different uppercase letters and lower case
 6 letters in the same bar is statistically significant (Duncan multiple comparison test, P<0,001).
 7

8 **Figure 6.** The effects of amino acid treatments on ABA content of broccoli
 9 under salinity. S0: 0 mM NaCl, S1: 100 mM NaCl, Ga: Gluten amin and Pr:
 10 Protein.



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 12 The difference between the means indicated by different uppercase letters and lower case
 13 letters in the same bar is statistically significant (Duncan multiple comparison test, P<0,001).
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1 **References**

- 2
- 3 Akram NA, Hafeez N, Farid-ul-Haq M, Ahmad A, Sadiq M, Ashraf M (2020). Foliage
4 application and seed priming with nitric oxide causes mitigation of salinity-
5 induced metabolic adversaries in broccoli (*Brassica oleracea* L.) plants. *Acta*
6 *Physiologiae Plantarum*, 42, 1-9.
- 7 Albacete A (2020). Get together: the interaction between melatonin and salicylic acid
8 as a strategy to improve plant stress tolerance. *Agronomy*, 10(10), 1486.
- 9 Ali Q, Haider M Z, Shahid S, Aslam N, Shehzad F, Naseem J, Ashraf R, Ali A,
10 Hussain S M (2019). Role of amino acids in improving abiotic stress tolerance to
11 plants. In *Plant tolerance to environmental stress* (pp. 175-204). CRC Press.
- 12 Ali L, Shaheen MR, Ihsan MZ, Masood S, Zubair M, Shehzad F (2022). Growth,
13 photosynthesis and antioxidant enzymes modulations in broccoli (*Brassica*
14 *oleracea* L. var. *italica*) under salinity stress. *South African Journal of*
15 *Botany*, 148, 104-111.
- 16 Angelini R, Federico R (1989). Histochemical evidence of polyamine oxidation and
17 generation of hydrogen- peroxide in the cell wall. *J.Plant Physiol.*, 135, 212-217.
- 18 Angelini R, Manes F, Federico R, (1990). Spatial an functional correlation between
19 daimine- oxidase and peroxidase activities and their dependence upon
20 deetilation and wounding in chick-pea. *Planta*, 182, 89–96.
- 21 Bahari A, Pirdashti H, Yaghubi M (2013). The effects of amino acid fertilizers
22 spraying on photosynthetic pigments and antioxidant enzymes of wheat (*Triticum*
23 *aestivum* L.) under salinity stress. *International Journal of Agronomy and Plant*
24 *Production*, 4(4), 787-793.
- 25 Bates LS, Waldren RP, Teare ID (1973). Rapid determination of free proline for
26 waterstress studies. *Plant Soil*, 39, 205-207.
- 27 Battal P, Tileklioglu B (2001). The effects of different mineral nutrients on the levels
28 of cytokinins in maize (*Zea mays* L.). *Turk. J. Bot.*, 25, 123-130.
- 29 Chevilly S, Dolz-Edo L, Morcillo L, Vilagrosa A, López-Nicolás J M, Yenush L,
30 Mulet J M (2021). Identification of distinctive physiological and molecular
31 responses to salt stress among tolerant and sensitive cultivars of broccoli
32 (*Brassica oleracea* var. *Italica*). *BMC Plant Biology*, 21(1), 1-16.
- 33 Danquah A, De Zélicourt A, Colcombet J, Hirt H (2014). The role of ABA and MAPK
34 signaling pathways in plant abiotic stress responses. *Biotechnology*
35 *advances*, 32(1), 40-52.
- 36 Haghighi M, Saadat S, Abbey L (2020). Effect of exogenous amino acids application
37 on growth and nutritional value of cabbage under drought stress. *Scientia*
38 *Horticulturae*, 272, 109561.
- 39 Hasanuzzaman M, Bhuyan MB, Zulfiqar F, Raza A, Mohsin S M, Mahmud JA, Fujita
40 M, Fotopoulos V (2020). Reactive oxygen species and antioxidant defense in
41 plants under abiotic stress: Revisiting the crucial role of a universal defense
42 regulator. *Antioxidants*, 9(8), 681.
- 43 Hasanuzzaman M, Raihan MRH, Masud A A C, Rahman K, Nowroz F, Rahman M,
44 Nahar K, Fujita M (2021). Regulation of reactive oxygen species and antioxidant
45 defense in plants under salinity. *International Journal of Molecular*
46 *Sciences*, 22(17), 9326.
- 47 Kang G, Li G, Guo T (2014). Molecular mechanism of salicylic acid-induced abiotic
48 stress tolerance in higher plants. *Acta Physiologiae Plantarum*, 36, 2287-2297.
- 49 Kumar A, Singh S, Gaurav AK, Srivastava S, Verma J P (2020). Plant growth-
50 promoting bacteria: biological tools for the mitigation of salinity stress in
51 plants. *Frontiers in Microbiology*, 11, 1216.

- 1 Kuraishi S, Tasaki K, Sakurai N, Sadatoku K (1991). Changes in levels of cytokinins
2 in etiolated squash seedlings after illumination. *Plant Cell Physiol.*, 32, 585–591.
- 3 Liu S, Dong Y, Xu L, Kong J (2014). Effects of foliar applications of nitric oxide and
4 salicylic acid on salt-induced changes in photosynthesis and antioxidative
5 metabolism of cotton seedlings. *Plant Growth Regul.*, 73, 67–78.
- 6 Matysiak K, Kierzek R, Siatkowski I, Kowalska J, Krawczyk R, Miziniak W (2020).
7 Effect of exogenous application of amino acids l-arginine and glycine on maize
8 under temperature stress. *Agronomy*, 10(6), 769.
- 9 Mushtaq Z, Faizan S, Gulzar B (2020). Salt stress, its impacts on plants and the
10 strategies plants are employing against it: A review. *Journal of Applied Biology
11 and Biotechnology*, 8(3), 81-91.
- 12 Peña Calzada K, Olivera Vicedo D, Habermann E, Calero Hurtado A, Lupino Gratão
13 P, De Mello Prado R, Lata-Tenesaca LF, Martinez CA, Celi GEA, Rodríguez JC
14 (2022). Exogenous application of amino acids mitigates the deleterious effects of
15 salt stress on soybean plants. *Agronomy*, 12(9), 2014.
- 16 Park H J, Kim W Y, Yun DJ (2016). A new insight of salt stress signaling in
17 plant. *Molecules and Cells*, 39(6), 447.
- 18 Rai VK (2002). Role of amino acids in plant responses to stresses. *Biologia
19 Plantarum*, 45(4), 481-487.
- 20 Raza A, Salehi H, Rahman M A, Zahid Z, Madadkar Haghjou M, Najafi-Kakavand S,
21 Charagh S, Osman HS, Albaqami M, Zhuang Y, Siddique KHM, Zhuang
22 W(2022). Plant hormones and neurotransmitter interactions mediate antioxidant
23 defenses under induced oxidative stress in plants. *Frontiers in Plant Science*, 13.
- 24 Sharma E, Sharma R, Borah P, Jain M, Khurana J P (2015). Emerging roles of auxin
25 in abiotic stress responses. *Elucidation of Abiotic Stress Signaling in Plants:
26 Functional Genomics Perspectives, Volume 1*, 299-328.
- 27 Sahin U, Ekinçi M, Ors S, Turan M, Yildiz S, Yildirim E (2018). Effects of individual
28 and combined effects of salinity and drought on physiological, nutritional and
29 biochemical properties of cabbage (*Brassica oleracea* var. capitata). *Scientia
30 Horticulturae*, 240, 196-204.
- 31 Turan M, Ekinçi M, Yıldırım E, Güneş A, Karagöz K, Kotan R, Dursun A (2014).
32 Plant growth-promoting rhizobacteria improved growth, nutrient, and hormone
33 content of cabbage (*Brassica oleracea*) seedlings. *Turk. J. Agric. For.*, 38, 327–
34 333.
- 35 Wang W, Cang L, Zhou D M, Yu Y C (2017). Exogenous amino acids increase
36 antioxidant enzyme activities and tolerance of rice seedlings to cadmium
37 stress. *Environmental Progress & Sustainable Energy*, 36(1), 155-161.
- 38 Wu X, Zhu W, Zhang H, Ding H, Zhang HJ (2011). Exogenous nitric oxide protects
39 against salt-induced oxidative stress in the leaves from two genotypes of tomato
40 (*Lycopersicon esculentum* Mill.). *Acta Physiol. Plant*, 33, 1199–1209.
- 41 Vaishnav D, Chowdhury P (2023). Types and Function of Phytohormone and Their
42 Role in Stress. IntechOpen. doi: 10.5772/intechopen.109325
- 43 Zhao S, Zhang Q, Liu M, Zhou H, Ma C, Wang P (2021). Regulation of plant
44 responses to salt stress. *International Journal of Molecular Sciences*, 22(9), 4609.
- 45 Zhu JK (2002). Salt and drought stress signal transduction in plants. *Annual Review of
46 Plant Biology*, 53(1), 247-273.
- 47